

off at Bathhurst Inlet, where all that I saw were a few battered *C. palæno*, five *P. aquilo* and one *L. phlæas feildeni*, even less marked than the Coppermine ones, but with only one specimen I cannot make any comments.

After two weeks at Bathhurst I finally went on to Victoria Island, and later to the Boothia Peninsula, but by that time the season was over and not a bug to be seen. The flowers were all in seed and the migratory birds already beginning to collect for their long flight south. At last an aircraft turned up and flew me out, but not until the first blizzard had come and the lakes begun to form ice. I had seen the short arctic summer come and go — it had been all too short for me. However, I had got pretty well everything I wanted, and a few things I had never expected to see in my life outside of a museum, so I was happy. Apart from that, there is a calmness and peace about the Arctic which is most soul-satisfying. I hung on in the North until the sea freeze-up started, and when I finally returned to the "outside" at Halifax by way of Baffin Island and Labrador, I swore I would return to the Arctic at the first opportunity.

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NOTES ON TECHNIQUES

Every collector has pet techniques or ideas. Some of mine may be of interest to other amateurs.

Unlike most entomologists, I use ethyl acetate as the killing agent in my jars. Among the advantages are: 1) It is non-poisonous to humans. Thus it can be safely used around children. 2) Even though the insect stops fluttering within a few seconds, it can be revived even some minutes after capture for release or egg-laying without ill effects. This is especially useful for a collector who rears. 3) Jars may be easily made by pouring plaster into wide-mouthed screw-cap jars as with other killing jars. When the plaster dries the ethyl acetate can be poured in as needed. 4) Specimens stiffen only upon very long contact with the reagent. One must be careful not to soak the plaster too much, or the insect may become wet and harden. The ethyl acetate evaporates from the specimen rapidly, leaving the scales unaffected. The stiffness remains, however, but can be eliminated by a short stay in a relaxing box.

Some disadvantages are as follows: 1) Many collectors want a killing agent that acts quickly. 2) Since the ethyl acetate is volatile, jars must be recharged frequently. 3) Rubber ringed caps can not be used, for the reagent softens and dissolves rubber. This can be avoided by inserting a tight cardboard disc into the cap. The cardboard can be covered with aluminum foil or wax to make a tight seal. Needless to say, to me the advantages outweigh the disadvantages.

For several years, I have covered my spreading boards with graph paper attached by means of soft glue. The squares speed mounting and enable more accurate positioning of the wings. The soft glue allows pins to go through very readily. When the boards get too worn, the paper can be sanded down and a new layer pasted on.

In rearing larger larvæ, I use wide-mouthed glass jars or large waxed cottage cheese cartons. These can be easily cleaned and washed. Netting can be fastened over the mouth of the container with a rubber band. On very humid days when moisture collects in the containers, they can be held up to a fan for a few minutes whenever needed.

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